# Matrix quality variability of oilseed radish (*Raphanus sativus* L. var. *oleiformis* Pers.) and features of its formation in technologically different construction of its agrophytocenosis

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**Abstract.** Overview of the formation of matrix variability of oilseed radish seeds at two levels: within a single pod and within the generative part of the plant, given the presence of vertical layering in the placement of fruit elements. The peculiarities of formation of variation component of morphological parameters of seeds from different zones of inflorescence were estimated, basing on their electrical scanning for oilseed radish agrophytoconosises of different technological construction and sowing rate against the background of four options of mineral nutrition (without fertilizer to the level 90 kg ha<sup>-1</sup> with an equal interval of 30 kg ha<sup>-1</sup> of the primary material). The results of the value of matrix variability in morphological and weight characteristics were grounded on the basis of the analysis of the structure of selected seed fractions and their intra-group variation. The main features of formation of morphometric variability of seeds from different zones of inflorescence depending on different technological construction.

**Key words:** heterospermy, matrix variability, morphological parameters of seeds, oilseed radish, seed fractions, separation variability.

### INTRODUCTION

Modern approaches to seed production technologies provide for the transition from general concepts of qualitative categories of seeds in general to the study of complex patterns of its formation. This provides prediction of its yield, sowing and adaptive properties and allows obtaining seeds with a high level of genetic (matrix), ecological-adaphic and morphological uniformity. Seed uniformity in this aspect is considered as a factor in ensuring optimal and uniform germination rates, which forms a high-intensity agrophytocenosis of the culture with the same standing density, the desired level of coenotic stress and feeding conditions (Scarisbrick, 1982; Wulff, 1986; Bouttier & Morgan 1992; Aicher, 2011; Balodis & Gaile, 2016; Stankowski et al., 2019; Wilczewski et al., 2020).

Heterospermia of seeds is considered as its difference in morphological characteristics, biochemical composition and physiological state, the ability to germinate and certain provide productivity of plants in offspring (Venable, 1985; Geritz et al., 1999; Alonso-Blanco et al., 2009; Li et al., 2014, 2014a; Yang et al., 2017).

Scientific practice distinguishes three types of heterospermy, namely ecological, matrix and genetic (genotypic) (Kizilova, 1974; Makrushin, 1994; Zhu & Weir, 1994; Yizhyk, 2000, 2000a; Rahman & Vetty, 2011; Hua et al., 2012). Ecological quality variability is the result of interaction of seeds in the process of its formation with environmental factors and belongs to not hereditary form, but it is an important aspect of the formation of biological and morphological properties of seeds and levels of seed productivity in the format of implementation of the genotype-environment combination (Leishman, 1995; Rees, 1996; Zhu, 1996; Fox et al., 1997; Nikolayeva, 1999; Imbert, 2002; Yang & Midmore, 2005; Gunasekera, 2006; Silveira et al., 2012; Sadras et al., 2013; Zhang et al., 2013; Ivanova & Sarmosova, 2014; Gnan et al., 2014; Yang et al., 2017; Li et al., 2020).

Genotypic variability is the result of a combination of hereditary signs of parental forms. In these conditions, while preserving the general type of heredity (varietal characteristics), each seed has differences due to the insemination process. This type of variability is also caused by mutagenic factors (Tayo & Morgan, 1978; Clarke, 1979; Morgan, 1980; Kindruk, 1990; Smith & Scarisbrick, 1990; Batygina, 1999; Ali et al., 2002; Wolfe & Mazer, 2005; Kaushik et al., 2007; Kennedy et al., 2011; Würschum et al., 2012; Li et al., 2014; Fu et al., 2015; Li & Li, 2015a; Li et al., 2015b; Li et al., 2019).

Matrix variability is one of the most common options and is formed as a result of different placement of flowers within the inflorescence on the mother plant. As a result, conditions are created for different nutrition conditions of different inflorescence levels and different influence of the plant as a mother body. Even if genetic and environmental factors are identical, the location of the seeds leads to the heterospermy detection (Olsson, 1960; Cavers & Steel, 1984; Inanaga & Kumura, 1987; Roach & Wulff, 1987; Smith & Scarisbrick, 1990; Habekotté, 1996; Makrushin, 1994; Donohue & Schmitt, 1998; Mazer & Wolfe, 1998; Nikolayeva, 1999; Sundaresan, 2005; Sadeghietal., 2011; Bukharov & Baleev, 2012; Hua et al., 2012; Zhang et al., 2016: Zheng et al., 2017).

The primary reasons for the matrix variability are the differences associated with the placement of individual seeds in different parts of the inflorescence in the vertical and horizontal orientation, namely: different flowering time, insemination and seed formation, different input intensity of plastic substances in the process of seed formation due to the reutilization of their transformation from the vegetative part of the mother plant fruits and seeds, as well as different degree of protection of the seedor fruit from abiotic environmental factors (Tayo & Morgan, 1978; McGregor, 1981; Roach & Wulff, 1987; Chay & Thurling, 1989; Kindruk, 1990, Makrushin, 1994; Donohue & Schmitt, 1998; Lemontey et al., 2000; Zlobin, 2009; Faraji, 2010; Kennedy et al., 2011; Wang et al., 2011; Gomez & Miralles, 2011; Gnan et al., 2014; Khan et al., 2014; Xing et al., 2014; Yang et al., 2016, 2017). This changes the physical and mechanical, chemical, sowing and yield properties of seeds. For these reasons, the creation of a control system for matrix heterospermia is a more important prerequisite for obtaining high-quality seed (Berry & Spink, 2006; Faraji, 2012; Monty et al., 2016).

As biological and physiological aspects, the matrix quality variability of seeds is important in the analysis of adaptability of technology and optimality of formation of agrophytocenosis both from the position of plant density in the unit area and from the position of mineral nutrition (Tsytsiura, 2019). For the group of cruciferous crops, among which is the object of our study, the signs of heterocarpy and heterospermy were studied by separate indicators in a number of studies. Thus, the results of the study of peculiarities of flowering stages of the generative part of rapeseed (McGregor, 1981; Habekotté, 1996.) and mustard (Vovchenko & Fursova, 2012) were made public. The peculiarities of forming the spatial structure of winter and spring rapeseed are highlighted (Tayo & Morgan, 1978; Smith & Scarisbrick, 1990; Habekotté, 1996). Issues of modeling the architectonics of rapeseed inflorescence and estimation of peculiarities of fruits and seeds uniformity within the inflorescence in such modeling are considered (Wang et al., 2011; Li et al., 2016). The influence of fractional composition of seeds of a number of cruciferous crops on the consistent formation of its sowing and yielding qualities was grounded (Olsson, 1960; Nikolaeva et al., 1999, Vyshnivsky, 2014). The evaluation of the general stage of flowering period of individual cruciferous crops and their influence on the formation of conditioned seeds was carried out (Smith & Scarisbrick, 1990; Polowick & Sawhney, 1986).

It is also noted for many cruciferous crops that the degree of variability of seeds is determined by the level of mineral nutrition of plants (Gattelmacher et al., 1994; Ozturk, 2010; Béreš et al., 2019), as well as technological factors such as the nature of the combination of fertilizer (Vujaković et al., 2014), the relationship between macro and micronutrients in the fertilizer system (Jankowski et al., 2014). In the formation of morphological features of seed development within the inflorescence a significant influence on this indicator of the technological features of nitrogen nutrition. In terms of flowering duration, the nature of flower formation, the use of phosphorus fertilizers and trace elements, as well as their ratio (Gomez & Miralles, 2011; Grant et al., 2013; Ragab et al., 2015; Sieling et al., 2017; Béreš et al., 2019; Zou et al., 2020).

The area of plant nutrition in the agrocoenosis due to the regulation of the value stress in turn determines the morphological development of plants, reproductive effort and seed productivity. The increase in standing density per unit area is directly proportional to the decrease in the generative part of plants with a decrease in the reaction rate interval in terms of morphological and weight characteristics of fruits and seeds (Hocking & Stapper, 2001; Takashima et al., 2013). As a result, the morphometric and weight alignment of seeds for many cruciferous crops, including oilseed radish, will be determined by the optimal ratio of technological parameters of subsowing formation of agrocenosis at the appropriate value of background mineral nutrition (Rathke et al., 2005; Al-Doori & Hasan, 2010; Qian et al., 2012; Sieling et al., 2017; Porter et al., 2020). For each plant species, these parameters are individual and rather extent unique (Berry et al., 2010; Qian et al., 2012). On the other hand, this conditions are different for different soil-climatic zones, but tend to be similar (Hocking et al., 1997; Horst et al., 2003; Marjanovi'c-Jeromel et al., 2011). It is also noted that considering the aspects of the matrix variability of seeds, given its fractional composition during harvesting and separation, provides a reduction of yield losses up to 30–39% and significantly improves the homogeneity of seed material and provides optimization of growth processes for agrophytocenosises basing on its consistent reproductive use (Makrushin, 1994; Kyrpa, 2011).

On the other hand, it was noted (Hasanuzzaman, 2020) that despite the agro-technological value of a number of crops from the cruciferous family. These factors are studied in detail on winter and spring rape and little-studied on white and black mustard, spring colza, and the object of our study oilseed radish as well.

For this group of crops, which also includes oilseed radish (Raphanus sativus var. *oleifera* Pers.), in addition to the above types of seed variability within its matrix type, its individual subtypes are distinguished: gravimorphic (differences in seed weight, size, shape, pattern) and enantiomorphic (differences in symmetry and asymmetry of a seed or fruit) (Dorofeyev, 2004; Tsytsiura & Tsytsiura, 2015). A number of questions remain to be discussed concerning the estimation of the general level of variation of morphological parameters of seeds in cruciferous crops and their influence on the value of realization of individual seed productivity of plants. It is also important to consider the peculiarities of layering in the flowering of different orders of the inflorescence of cruciferous crops (Schiessl et al., 2015) and, as a consequence, the corresponding stage in the formation of fruits from different tiers in the vertical placement of the last from the base of the inflorescence to its apex. This feature is noted by a number of researchers on winter and spring rapeseed (McGregor, 1981; Scarisbrick et al., 1982; Habekotté, 1996; Wang et al., 2011; Vyshnivsky & Slisarchuk, 2014) but is poorly studied on oilseed radish plants. This leads to well-known technological problems in the time diversity of seed maturation and choice of pre-harvest preparation of plants, associated with the need for separate harvesting or the use of desiccation, the latter has certain limitations in the application for the oilseed radish agrophytocenosises (Tsytsiura, 2018a). Given the above facts, the assessment of the level of matrix variability of oilseed radish varieties will make it possible to determine the adaptation strategy of plants, and use this parameter as an indicator of the optimization of agrophytocenosis formation.

### **MATERIALS AND METHODS**

The research on the formation of indicators of the matrix variability of oilseed radish varieties of plants with changes in technological parameters of the construction

of its agrophytocenosises was carried out during 2013–2018 on the experimental field of theVinnytsia National Agrarian University (N 49°11'31", E 28°22'16") on dark gray forest soils (Luvic Greyic Phaeozem soils (Working Group WRB, 2015)) of the medium loam mechanical compositi on with fluctuating basic agrochemical indicators in the rotation section: humus content 2.16–2.52%, pH 5.8–6.7, lightly hydrolyzed nitrogen content 71–77 mg kg<sup>-1</sup>, mobile phosphorus 187–251 mg kg<sup>-1</sup>, exchangeable potassium 95–143 mg kg<sup>-1</sup>. The basic technological scheme of technological construction of

**Table 1.** The range of acceptable common options for the formation of oilseed radish agrophytocenosis in the study area (Tsytsiura, 2019)

Planting met	Fertilization (of the active	
rates (million gern	substance), kg ha <sup>-1</sup>	
Row method	Wide-row method	Without
(15 cm)	(30 cm)	fertilizers
1.0	0.5	$N_{30}P_{30}K_{30}$
2.0	1.0	$N_{60}P_{60}K_{60}$
3.0	1.5	N90P90K90
4.0	2.0	

oilseed radish agrophytocenosises provided for the application of the interval of technological solutions adopted in the region in terms of row width and sowing rate in

accordance with zonal recommendations of oilseed radish cultivation in the zone of theforest-steppe of moderately continental belt (Table 1). Term of sowing of all variants corresponded to the period of the end of the first-middle of the second ten-days period of April.

Three recognized varieties of oilseed radish, namely, 'Zhuravka', 'Raiduha' and 'Lybid', were used in the research. These varieties belong to the varieties of combined use for fodder purposes and seeds with wide adaptive potential for cultivation in different soil and climatic zones.

The hydrothermal regime of oilseed radish vegetation during the period of seed formation and filling differed (Table 2).

0	8		1					
		Years						Long-time
Months	Ten-days							average annual
WIOIIUIS	period	2013	2014	2015	2016	2017	2018	value (30-year
								averaging period)
		Averag	e daily to	emperatu	re, °C			
	Ι	17.2	18.1	20.4	15.9	18.0	19.2	15.9
June	II	19.9	16.3	19.2	18.7	18.1	20.7	16.7
	III	20.8	15.7	18.2	23.7	21.4	17.9	17.5
	Ι	19.7	19.2	21.5	19.3	18.2	18.6	18.2
July	II	18.6	19.7	19.0	20.6	19.2	19.1	18.8
•	III	18.6	21.3	22.7	21.6	21.7	21.5	19.0
Average f	for the selecte							
period, °C		19.5	18.5	19.7	20.8	19.8	19.1	17.7
1		Precipi	tation an	ount, mi				
	Ι	36.0	43.1	3.2	15.0	1.8	0.5	22.8
June	II	71.0	0.0	28.8	38.9	11.9	91.3	24.7
	III	37.0	33.4	9.3	18.3	15.2	156.3	25.9
	Ι	0.2	28.5	3.0	17.4	5.7	10.0	25.2
July	II	11.1	39.0	11.9	44.0	18.6	44.0	23.8
2	III	15.4	15.0	5.8	6.7	67.7	52.3	29.2
Amount f	forthe selected							
period, m	m	134.7	216.8	56.2	125.3	119.1	302.1	151.6
<b>1</b>		HTC**						
May		1.305	2.783	0.719	1.227	0.645	0.258	1.460
June		2.202	1.078	0.613	0.893	0.349	3.124	1.416
July		0.377	1.137	0.230	0.682	0.806	1.349	1.396
August		1.047	0.750	0.061	0.486	0.563	0.349	1.234
Average f	for the	1.232	1.437	0.406	0.822	0.591	1.270	1.377
vegetation								
		. 1.0	a .	(DD GI	I (0) 1		1	

**Table 2.** Precipitation amount and average daily temperature in comparison with the long-time average annual regime for the research period

\* – grey color marks the period from flowering (BBCH 63) to the yellow-brown pod phase (BBCH 86) of the oilseed radish for phenological conditions of the study area; \*\*  $HTC = \frac{\Sigma R}{0.1 \cdot \Sigma t_{>10}}$ , where the amount of precipitation ( $\Sigma R$ ) in mm over a period with temperatures above 10 °C, the sum of effective temperatures ( $\Sigma t_{>10}$ ) over the same period, decreased by a factor of 10 (Polyovy & Bozhko, 2007).

Variation of morphological parameters of seeds was estimated with a sample of 25 plants in non-contiguous repetitions (total sample of 50 plants). Total number of repetitions for each variant 4. Plant analysis provided for the evaluation of a group of 5 plants in 5 places by the length of the line stochastically by the width of the area with a displacement in the line horizontal to the phase of brown maturity of the pod BBCH 87 (Hocking, 1997; Meier, 2001; Skinner & Moore, 2007) when all the pods have reached a typical size. The formed seeds were selected separately from the three zones of the oilseed radish inflorescence (evenly with the same interval on the total height for each inflorescence according to the general recommendations for determining the vertical level and mosaic of agrophytocenosises (Rabotnov, 1992)). Peculiarities of oilseed radish fruit anatomy due to the evaluation of heterospermy level were carried out taking into account scientific developments in the field of carpology for cruciferous crops (Puri, 1941; Zohary, 1948; Levina, 1987; Lotova & Rudko, 1999; Dorofeyev, 2004; Ilyinska, 2013.

Seed mass characteristics were determined with the use of laboratory scales RADWAG PS 1000.R1with 0.001 g discretion.Analysis of fractional composition of seeds was carried out by sifting the selected average sample of seeds on testing sieve sets with a round cell according to Specifications 23.2.2068-94 typeIandwitha rectangular cell according to Specifications 5.897–11722–95, TC 23.2.2068-94 type II. The fractional composition was also determined by the method of processing a single-layer scanned image of seeds using the Digimizer image analysis software package (v 4.2). Weight of 1000 seeds was determined according to the state national standard (State standard of Ukraine 4138-2002, 2003).

Seed parameters were estimated based on such characteristics as projection surface area (PS, mm<sup>2</sup>), surface perimeter (P, mm), seed length (L, mm), seed width (W, mm), roundness (R, mm), seed sorting index (SI) (ratio of seed length (L, mm) to its width (W, mm)).

Study of the morphological structure of fruits was conducted in accordance with the general recommendations of morphological and anatomical analysis of cruciferous fruits (Puri, 1941; Zohary, 1948; Levina, 1987; Lotova & Rudko, 1999; Dorofeyev, 2004; Ilyinska, 2013).

To determine these morphometric characteristics, used Didital Caliper (measurement accuracy of 0.01 mm), USB microscopy SigetaMCMOS 5100 5.1 MPUSB 2.0 (in combination with a digital microscope for  $\times 10$  and  $\times 40$  optical magnification formats) and OotdtyDM–1600, 2 MP with corresponding software and using ImageJ 1.52 software.

The matrix variability index (MVI) in comparison with the morphological parameters of seeds from the lower and upper inflorescence zone was determined in relation to the average coefficient of variation (CV,%). Measurements and observations were performed using field and laboratory photography with the camera Canon EOS 750DKit and lenses Canon EF 100 mmf/2.8 LUS Mand CanonMP-E 65 mmf/2.8 1-5 x Macro.

General methodology of researches, consideration of phenological phases in the system of signs of oilseed radish bearing phase and other related observations and records were conducted in accordance with the basic recommendations of researches on cruciferous crops (Sayko, 2011; Test Guidelines..., 2017) using the package of statistical software Statistica 10, Exel 2013, Past 324 and biometric methods of statistics

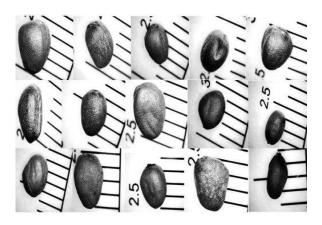
(Zar, 1984). The level of variability of morphological features and grouped indicators was conducted on the scale (Zaytsev, 1984): very low (CV < 7%); low (CV = 8-12%); medium (CV = 13-20%); elevated (CV = 21-30%); high (CV = 31-40%); very high (CV > 40%). Given the massiveness of the data and general similarity in the values of formation of features for the studied oilseed radish varieties. The data are presented by key indicators for 'Zhuravka' variety.

## **RESULTS AND DISCUSSION**

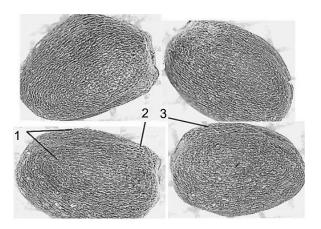
According to our preliminary estimates (Tsytsiura, 2018b), there is a great variation in the seeds of oilseed radish varieties. The seed of this crop itself has a number of

morphological forms, which, in average, make it possible to attribute it to the egg-shaped one with different index of length to width ratio. This is clearly demonstrated in Fig. 1.

According to the presented typological forms of seeds there are egg-shaped, elongated egg-shaped, flattened egg-shaped, oval, globeshaped and various combined forms. Seed color is also a variable indicator (gravimorphic variability) from grey and brown to dark brown with a dark red shade. The color tends to change from light brown for newly harvested seeds to brownish red for seeds after long-term storage. Common external morphological features can be identified for any oilseed radish seed of any shape (Fig. 2). The seed is divided into two symmetrically cotyledonary lobes, separated by a shallow hollow of hypocotyl and coleorhiza elements. These elements are most noticeable on seeds of the middle zone of the pod. The formation of seeds of different sizes within the pod for oilseed radish is determined by several reasons. Some of them are associated with the peculiarities of the formation of different morphotypes

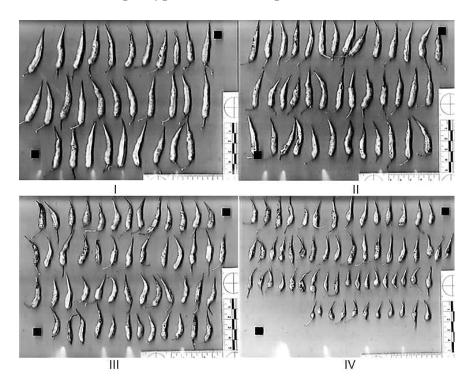


**Figure 1.** Quality variability of oilseed radish seeds of the 'Zhuravka' variety by typological linear sizes and shape, 2015.



**Figure 2.** General appearance of oilseed radish seeds of the 'Zhuravka' variety from different positions, 2018.

of pods in the context of different zones of inflorescence, determined by a certain layering of formation of fruit elements the gradient of height (Tsytsiura & Tsytsiura, 2015; Tsytsiura, 2018, 2019). Depending on the tier (lower, middle and upper) (Fig. 3) the oilseed radish pods differ in length, diameter and their ratioas well as by the wall thickness of the pod itself. In particular, the pods of the upper zone in comparison with the pods of the lower zone have thicker walls, significantly shorter length of the pod itself and overall higher variability in its morphology. The staging of the phenological formation of pods of the upper zone also has signs of desynchronization in the format of accelerated formation and maturation. It causes to deformation of the fruit walls, and to the appearance of such morphotypes as twisted, spiral, crescent, etc.



**Figure 3.** Morphotypes of oil seed radish pods of the 'Zhuravka' variety (I – pods of the lower zone; II – pods of the middlezone; III, IV – pods of the upper zone) for thetechnological variant of 2.0 mlnpcs. ha<sup>-1</sup> of the germinable seeds with wide-row sowing against the back ground  $N_{90}P_{90}K_{90}$  (for the phenological phase BBCH 87–88) (black square with dimensions 2×2 cm), 2018.

This nature of change in the morphology of the fruit provides not only a general decrease in the number of seeds in the pod in the direction from the base to the apex of the inflorescence at the oil radish. This is the scope of variation in the shape and size of the seed and its individual weight characteristics. Such features of pod formation for oilseed radish create prerequisites for reducing the favorability of the formation of the seed itself. This eventually leads to a change in the shape of the seed itself. Similar features were noted on a number of agricultural crops, including those of the cruciferous family (Levina, 1987; Pechan, 1988; Bouttier & Morgan, 1992; Mazer & Wolfe, 1998; Faraji, 2010, Hua et al., 2012; Gnan et al., 2014; Li et al., 2014, Li & Li, 2015a; Li et al., 2015b; Yang et al., 2016, Li et al., 2019, 2020).

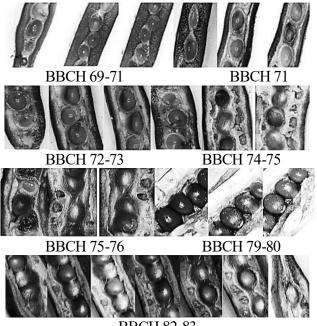
The matrix variability of seeds in oilseeded radish pods within one inflorescence zone is also due to the peculiarities of its formation from the earliest stages after successful insemination. Conducted anatomical and microscopic studies at the stage of initiation of the formation of oilseed radish pods (BBCH 68-71), which indicate marked differences both in the size of the seed at the beginning of its formation and in the staging of its development. With these features, the format of the matrix variability by the height placement factor of a seed in a pod can be defined as differently directed. Over the period of many years of observations, I have found a gradual decrease in linear sizes of seeds

from the base of the pod to its apex and a high probability of seeds formation with changed morphotype in the lowest parts of the pod. It should be noted that for all zones

of oil radish inflorescence, which were determined during the evaluation of fruit elements by morphotype (Fig. 3), similar peculiarities in seed formation were noted. Such features are clearly shown in Figs. 4–5.

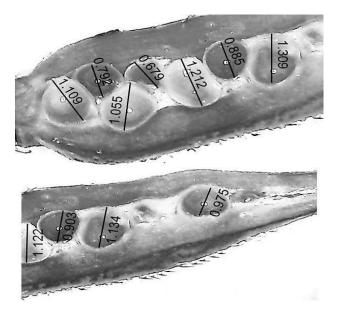
As a result, for the period of waxy ripeness of seeds (brown pod phase BBCH 85-87) in the dynamic row of assessment of linear sizes of seeds, placed similarly to the pod from its base to the top, there is a gradual reduction of seeds in sizes and the appearance of atypical seeds in shape in the row of seeds by pod height. At the same time, the total number of seeds in the pod has an overall stable tendency to decrease. It was 1-3 seeds on average in comparison with the lower zone pods to the upper zone pods. This is clearly illustrated in Fig. 6. It should be noted that in contrast to rapeseed and mustard, for which the general nature of the variation of linear seed size is most significant in the upper and lower pod zones (Clarke, 1979; Smith & Scarisbrick, 1990; Bouttier & Morgan, 1992; Angadi et al., 2000; Garcia et al., 2005; Gunasekera et al., 2006; Vovchenko & Fursova, 2012; Zheng et al., 2017; Luo et al., 2018; Li et al., 2019), for oilseed radish such variability is more pronounced zonally and includes some seeds of the middle zone of the pod.

The data array obtained in the automated scanned images processing mode (Table 3) has confirmed the general conclusion about the presence of significant levels of matrix quality variability of seeds. In



BBCH 82-83

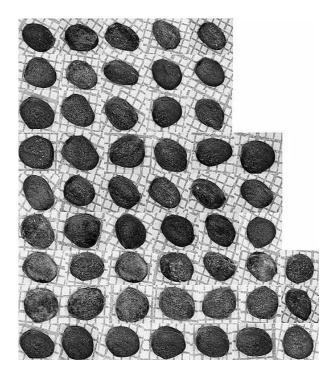
**Figure 4.** Nature of variability of oilseed radish seed size of the 'Zhuravka' variety in a pod at various microstages of its formation (in each line position of the figure from left to right: from the base of the pod to its apex), 2017.



**Figure 5.** Nature of variability of oilseed radish seed size of the 'Zhuravka' variety in a pod at various microstages of its formation position – seed size difference in the formation phase (BBCH 69-71, the dimension of values of mm), 2017.

comparison with seeds from pods with different placement on the mother plant, respectively, for the lower and upper zones against the background of increased variation (according to the values CV (Zaytsev, 1984) projection surface area ofseeds (SA), medium and elevated levels of seed surface perimeter variation (P), medium variation level in length (L), width (W) and seed roundness (R) and, respectively, elevated variation level in seed sorting index (SI) – variation coefficient value was 0.7–3.6% higher for seeds from the upper zone pods.

It was also determined that the studied technological parameters influenced the value of variation of the specified morphological features of seeds. Thus, for all the stand densities study, mineral under fertilizer application contributed to the growth of the overall overall value of all the features without exception, with increasing dynamics, in line with the increasing rate of fertilizer application. This nature of influence is more pronounced in the formation of seeds from the pods of the upper zone with an increase to the unfertilized variant the context of the studied in technological variants of sowing rates from 1.3 to 2.4% in comparison to seeds from the pods of the lower zone in the range from 1.0 to 1.9%. The gradual increase in variation of morphological signs of seeds with a decrease in plant stand density and an increase in the area of their nutrition with a maximum expression at the application of fertilizers in the norm of 90 kg ha<sup>-1</sup> of the primary material are determined. Thus, the difference in the average variation coefficient ( $CV_{aver.}, \%$ )



**Figure 6.** Qualityvariability of oilseed radish seeds of the 'Zhuravka' variety in the order of their placement in the pods of various zones of the generative part (three upper positions - pods of the upper zone, three middle positions - pods of the middle zone, three lower positions - pods of the lower zone; placement of seeds on the rows of the figure from left to right in each row from the base of the pod to its apex), sampling selection of 2017.

between the variant with the maximum stand density at the rate of 4.0 million of germinable seeds  $ha^{-1}$  with the fertilizer  $N_{90}P_{90}K_{90}$  and the variant with the minimum stand density at the rate of 0.5 million of germinable seeds  $ha^{-1}$  was 7.2% for seeds from the pods of the lower zone and 10.1% for seeds from the pods of the upper zone.

As a result, according to the matrix variability index (MVI), the most significant gap in the morphology of seeds of pods of the lower and upper zones with the value of 0.86 was noted exactly for the variant of 0.5 million of germinables seeds ha<sup>-1</sup> with the maximum fertilization with N<sub>90</sub>P<sub>90</sub>K<sub>90</sub>. Similar features are noted in other cruciferous crops (Kitaeva, 1952; Tayo & Morgan, 1978; Morgan, 1980; McGregor, 1981; Voytenko, 1989; Lovett-Doust, 1989; Rao et al., 1992; Habekotté, 1996; Susko & Lovett-Doust, 2000; Faraji, 2010; Wang et al., 2011; Li et al., 2016, 2019, 2020).

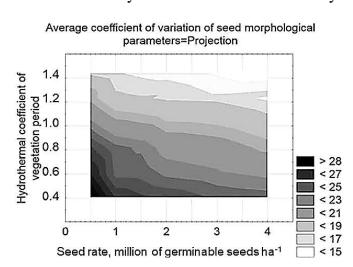
**Table 3.** Variation coefficients (CV, %) of the main signs of oilseed radish seeds of the 'Zhuravka' variety according to the results of scanning of seeds for the brown pod phase (BBCH 85-86) in different stages of inflorescence at different variants of formation of agrophytocenosis on the average for 2013-2018, % (for *N* in the scanning group from 4,200 to 18,000)

		Morphological signs of seeds that were observed in the Digimizer image analysis software (v 4.2)														
Sowingrate and sowing method	Ţ	PS, mm <sup>2</sup>	P, mm	L, mm	W, mm	R, mm		V <sub>aver</sub> , %	PS, mm	P, mm	L, mm	W, mm	R, mm		V <sub>aver</sub> , %	
ngn wow bo	Fertilizer						SI	CV	$\mathbf{PS}$	Ρ,	L,	M	R,	SI	CV	_
Sowing and sov method	artil	Seed	ls froi	m the	pods	of the	e upp	er	Seed	s from	n the	pods	of the	e low	er	N
		<u> </u>		e zone							e zone					Σ
4.0 million,	1			15.2												
row	2			15.8												
	3			16.4												
	4			17.2												
3.0 million,	1			16.2												
row	2			17.5												
	3			18.2												
	4			18.7												
2.0 million,	1			19.5												
row	2			19.8												
	3			20.2												
	4			20.7												
1.0 million,				20.8												
row	2			21.5												
	3			22.3												
	4			22.6												
2.0 million,				15.7												
wide-row	2			16.4												
	3			16.8												
	4			17.4												
1.5 million,				17.8												
wide-row	2		-	18.4			-			-						
	3			19.2												
	4			19.6												
1.0 million,				20.6												
wide-row	2			21.4												
	3			22.5												
<u> </u>	4			22.9												
0.5 million,				22.8												
wide-row	2			23.5												
	3			24.1												
	4	33.1	32.2	25.6	21.8	25.7	22.6	26.8	30.1	28.7	20.9	17.9	22.7	18.3	23.1	0.86
Tukey test upper/lowe zone <i>p<sub>adj</sub></i>	er	0.00096**	$0.04152^{*}$	$0.06317^{*}$	$0.03942^{**}$	$0.04752^{*}$										
												-	0 0 <b>-</b>			

 $1 - \text{Fertilizer-free}; 2 - N_{30}P_{30}K_{30}; 3 - N_{60}P_{60}K_{60}; 4 - N_{90}P_{90}K_{90}. \text{ Tukey test: } * - P < 0.05; * -P < 0.001.$ 

In my opinion, the obtained values are determined by the peculiarities of relations between plants in populations of different densities. Thus, at maximum technological density, the level of variation of indicators is under much higher coenotic tension than in the variants of minimum technological density. Due to these regularities, the average value of a feature has a smaller scale of deviations, also in terms of morphometry of seeds and fruit elements. Mineral fertilizers should be considered in our system as stress regulators, which reduce the coenotic tension (Zlobin, 2009). For these reasons, the range of deviations from the average for the same density level of cenosis indicated by

us also increases dynamically with a reduction of the coenotic tension lower sowing rates. at An important aspect in this context is determine to the role of hydrothermal conditions of oilseed radish vegetation on the general manifestation of the level of variation of morphological features of seeds, which was noted by a number of studies conducted on rapeseed (Rao et al., 1992: Morrison & Stewart, 2002; Gan et al., 2004; Lester et al., 2004; Luo et al., 2018). Comparison of factors of such influence (Fig. 7) allowed noting that decrease of hydrothermal coefficient (HTC) of oilseed radish vegetation period contributes to growth of general variability of seeds and expression of the effect of its matrix variability.



**Figure 7.** Dependence between the level of variation in the morphological parameters of oilseed radish seeds of the 'Zhuravka' variety, hydrothermal regime of the vegetation period and sowing rate of million germinable seeds ha<sup>-1</sup> in conjunction with the width of the row spacing for the average for the fertilizer variants (consolidated data for the 2013–2018 period).

The maximum level of variation variability of morphological features of seeds at the level of CV = 27-30%, obtained at HTC in the range of 0.400–0.800 at sowing rate of 0.5–1.0 million germinable seeds ha<sup>-1</sup>. Accordingly, the minimum level of variability of morphological features of seeds CV = 12-15% - when combined with the following parameters: HTC 1.3–1.4, 3.0–4.0 million germinable seeds ha<sup>-1</sup>. Such results are quite consistent with our conclusions about the role of stress levels in the combination of density growth. An important component of the evidence of the above conclusions is the conducted fraction separation of oilseed radish seeds on the basis of the studied technological parameters of the construction of its agrophytocenosises. The results of seed separation on two types of sieves with round holes ((mm): 0.5; 1.0; 1.5; 2.0; 2.5; 3.0; 3.5; 4.0; 4.5; 5.0; 6.0; 7.0) (Table 4) and with rectangular holes (1.0×20; 1.2×20; 1.5×20; 1.7×20; 2.0×20; 2.2×20; 2.5×20; 3.0×20) (Table 5) allow us to generalize the peculiarities of the matrix variability of oilseed radish seeds.

**Table 4.** Fraction composition of oilseed radish seeds by its separation on sieves with round holes for the 'Zhuravka' variety on the brown pod phase (BBCH 85-87) in different tiers of inflorescence with different variants of agrophytocenosis formation on the average for 2013–2018,% (A factor - year)

		Fractions by seed separationon sieves with rectangular holes, mm (%)															
Sowingrate (C), sowing method (B)	Fertilizer (D)**	0.5	0.5	0.5	0.5	0.5	0.5	0.5	m <sub>1,000</sub> , g	0.5	1.0	1.5	2.0	2.5	3.0	3.5	m1,000, g
Sowi sowii (B)	Fertil	Seed gene			-	s of t	he up	oper			ds fro erative		-	s of t	he lo	wer	
1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
4.0	1	7.2	20.6	26.3	25.8	20.1	0.0	0.0	7.6	3.8	14.2	17.3	30.8	33.9	0.0	0.0	8.3
million,	2	6.6		27.2				0.0	7.7	3.2			31.6			0.0	8.5
row	3	6.2		28.4				0.0	8.2	2.1			33.2			0.0	8.8
	4	5.4		29.2				0.0	8.4	1.8			34.4			0.0	8.9
3.0	1	6.4		28.4				0.0	8.0	2.6			31.6			0.0	8.6
million,	2	5.6		29.1				0.0	8.3	1.6			32.7			0.0	8,9
row	3	4.9		29.5				0.0	8.7	0.9			33.1			0.0	9.3
2.0	4	4.6		30.2				0.0	9.0	0.6			33.8			0.0	9.5
2.0	1	5.3		29.3				0.0	9.2	1.1			31.4			0.0	9.7
million,	2	4.5		29.6				$\begin{array}{c} 0.0\\ 0.0\end{array}$	9.4	1.0 0.4			32.9			0.0 0.0	10.0 10.3
row	3 4	3.4 2.7		30.2 30.8				0.0	9.6 9.8	0.4	10.5 8.3		32.6 32.4			0.0	10.5
1.0	<del>4</del> 1	3.7		30.6				0.0	9.8 9.7	0.4			32.4			0.0	10.0
million,	2	3.5		31.2				0.0	10.0		9.5	-	31.8			0.0	10.0
row	$\frac{2}{3}$	2.7		31.6				0.0	10.0		7.8		31.2			0.0	10.4
10 **	4	1.9		32.2				0.0	10.1		5.7		31.6			0.0	10.0
2.0	1	6.1		26.9				0.0	8.2	2.1			33.4			0.0	8.7
million,	2	5.8		27.4				0.0	8.5	1.8			34.8			0.0	9.0
wide-row	3	4.8		28.5				0.0	8.8	0.8	9.2		36.5			0.0	9.2
	4	4.5		29.2				0.0	9.0	0.5	7.8	19.7	37.1	34.9	0.0	0.0	9.4
1.5	1	5.2	17.3	27.4	28.5	21.6	0.0	0.0	9.0	1.2	12.3	19.2	30.5	36.8	0.0	0.0	9.4
million,	2	4.8	16.7	28.1	29.1	21.3	0.0	0.0	9.2	0.8	11	19.6	30.4	38.2	0.0	0.0	9.6
wide-row	3	4.4	16.4	28.9	29.4	20.9	0.0	0.0	9.3	0.6	9.5	19.9	29.8	40.2	0.0	0.0	10.0
	4	3.9	15.9	29.5	29.8	20.9	0.0	0.0	9.7	0.4	6.8	20.6	29.5	42.7	0.0	0.0	10.2
1.0	1	4.5	15.2	29.2	23.3	27.8	0.0	0.0	9.5	0.0			25.7			0.0	10.2
million,	2	3.8		31.3				0.0					25.1			0.0	10.5
wide-row	3	3.5		32.6				0.0		0.0	7.2		24.3			0.0	11.0
	4	2.7		33.4				0.0	10.4		5.7		23.5			0.0	11.3
0.5	1	2.3		31.5				0.0	10.4				19.5			0.0	11.2
million,	2	1.9		32.7				0.0	10.8		7.8		19.3			0.0	11.5
wide-row	3	1.7		33.4				0.0	11.0		5.5		19.2			0.3	11.7
	4	1.5		34.3				0.0	11.1				19.4			0.3	12.0
$LSD_{05}A^*$		0.08							0.14				0.29				0.15
$LSD_{05}B$		0.05							0.08				0.16				0.09
$LSD_{05}C$		0.07							0.12				0.23				0.13
$LSD_{05}D$		0.07							0.12				0.23				0.13
$LSD_{05}AB$		0.12							0.20				0.40				0.22
$LSD_{05}AC$ $LSD_{05}AD$		0.17 0.17							0.29 0.29				0.57 0.57				0.31 0.31
$LSD_{05}AD$ $LSD_{05}BC$		0.17							0.29				0.37				0.31
LSD05DC		0.10	0.33	0.21	0.40	0.43			0.1/		0.20	0.30	0.33	0.70			0.10

Table 4 (continued)

1	3 4	4 5	6	7	8	9	10	11	12	13	14	15	16	17	18
$LSD_{05}BD$	0.10 0	0.35 0.21	0.48	0.43			0.17		0.20	0.36	0.33	0.70			0.18
$LSD_{05}CD$	0.14 0	0.50 0.29	0.68	0.61			0.23		0.29	0.51	0.47	0.99			0.25
$LSD_{05}ABC$	0.24 0	0.86 0.51	1.19	1.06			0.41		0.50	0.88	0.81	1.71			0.44
$LSD_{05}ABD$	0.24 0	0.86 0.51	1.19	1.06			0.41		0.50	0.88	0.81	1.71			0.44
$LSD_{05}ACD$	0.34 1	1.22 0.72	1.68	1.50			0.57		0.71	1.25	1.14	2.42			0.62
$LSD_{05}BCD$	0.20 0	0.70 0.41	0.97	0.87			0.33		0.41	0.72	0.66	1.40			0.36
LSD <sub>05</sub> ABCD	0.48 1	1.73 1.01	2.37	2.13			0.81		1.00	1.77	1.62	3.42			0.87
Share of															
influence A															
В		86.8764.61					56.4		17.12	2 67.90	) 59.06	543.74	ŀ		56.8
С	6.38 1	0.54 1.19	12.55	523.26	5		5.00		10.25	6.50	6.35	14.39	)		5.54
D	64.18 4	40.750.99	17.75	537.1	l		31.7		21.50	) 20.90	)28.37	7 23.38	3		30.8
AB	15.15 9	9.88 1.89	2.55	0.20			4.99		47.41	3.40	3.61	5.48			5.24
AC	0.10 0	).23 9.52	0.26	0.28			0.07		0.18	0.09	0.20	0.16			0.08
AD	0.91 0	0.65 7.87	0.26	0.59			0.56		0.30	0.43	0.52	0.41			0.54
BC	0.18 0	0.12 0.34	0.12	0.03			0.19		0.59	0.17	0.09	0.18			0.19
BD	0.88 0	0.13 1.28	23.64	48.18			0.48		1.09	0.32	0.53	11.29	)		0.46
CD	0.46 0	0.05 0.22	0.33	0.52			0.02		0.33	0.02	0.01	0.03			0.01
ABC	0.63 0	0.40 0.24	0.93	0.82			0.17		0.34	0.05	0.35	0.35			0.13
ABD	0.02 0	0.05 9.24	0.51	0.10			0.10		0.05	0.08	0.12	0.12			0.07
ACD	0.01 0	0.00 0.36	0.01	0.01			0.00		0.01	0.00	0.07	0.00			0.00
BCD	0.01 0	0.01 1.01	0.04	0.02			0.02		0.01	0.03	0.18	0.02			0.02
	0.76 0	0.31 0.12	0.67	2.15			0.13		0.78	0.08	0.33	0.41			0.04
	0.02 0	0.02 1.11	0.02	0.04			0.02		0.02	0.03	0.21	0.02			0.02

\* – in the system of value expression at deprivation of values,%; \*\* – 1 – Fertilizer-free; 2 –  $N_{30}P_{30}K_{30}$ ; 3 –  $N_{60}P_{60}K_{60}$ ; 4 –  $N_{90}P_{90}K_{90}$ .

According to the data presented in both tables morphological row of oilseed radish seeds of the 'Zhuravka' variety is placed in the range of 0.5-3.5 mm for the sieves with round holes and  $1 \times 20 - 3 \times 20$  for the sieves with rectangular holes. This confirmed the affiliation of seeds to the general group a rounded and flattened. It also allowed recommending requires the introduction of a system of combined sieves for technological cleaning and calibration of oilseed radish seeds in production conditions. In addition, the array of data obtained indicates that the fraction composition of the size of seeds from pods of different zones of the plant is significantly different according to certain difference indices  $LSD_{05}$  for the main factors and their interaction. Thus, when separating seeds on round sieves (Table 4), given the significance of the difference between the studied variants, it should be noted a reliable increase in the percentage of fine fraction 0.5 mm in the seeds from the pods of the upper zone in the context of all the variants of standing density and fertilization. In fact, for variants 0.5-1.0 million germinable seeds ha<sup>-1</sup> of this pod zone, this fraction was absent. As well as it is practically absent the fraction 3.0 and 3.5 mm for all technological variants. The distribution between the seed fractions in the range of 1.5–2.5 mm for seeds of the lower and upper tier of pods also differed significantly. Thus, on the average for the studied technological variants the seed fraction share of 2.0 mm and 2.5 mm for seeds from the pods of the lower zone was 29.83% and 39.40%, which is 11.8% and 69.0% higher than for seeds from the pods of the upper zone. At the same time, the actual nature of fractional distribution in all variants differed: the sum of fractions' share of 0.5-1.5 mm for seeds from pods of the lower zone averaged 30.57%, which is 19.33% lower than for seeds from pods of the upper zone. Seeds of 3.0 and 3.5 mm fractions in the research variants are stably noted for technological variants of 0.5 and 4.0 million germinable seeds ha<sup>-1</sup> in the variant of  $N_{60}P_{60}K_{60}$  and  $N_{90}P_{90}K_{90}$  application. The role of mineral fertilizers in changing fractional composition of oilseed radish seeds also had its own peculiarities. Thus, for the variants of evaluation of fractional composition of seeds from the pods of the upper zone, the dynamic growth of fertilizer rates provided a decrease in the share of 0.5 and 1.0 mm seed fractions with adequate growth of other fractions. For seeds from the pods of the lower zone, the effect of mineral fertilizers was of two-level direction: a decrease in the share of the 0.5 mm fraction, 1.0 mm with the growth of other fractions. In case of wide-row sowing variants assuming the sowing rate of 1.5 million germinable seeds ha<sup>-1</sup>, a decrease in the share of 2.0 mm seed fraction with an increase in the intensity of 2.5 mm share growth by 8.9–10.3% was noted. This indicates that on more dissolved sowings the character of formation of fractional composition of oilseed radish seeds has a more complex interfactional nature with the allocation of two or three main fractions. It should also be noted that the general regularity in the formation of fractional composition on the round sieves, despite the peculiarities in the fertilization section of specific technological variants, has its own characteristics for seeds from pods of both zones. Under these conditions, it is established that a stable growth of the share of large fractions of 2.0, 2.5 mm with a corresponding decrease of fractions of 0.5-1.0 mm. For certain technological variants, the impact of fertilizers on the proportion of the respective fractions had certain features, particularly for seeds from the pods of the upper zone for variants of 3.0–4.0 million germinable seeds ha<sup>-1</sup> with normal row sowing and 1.5–2.0 million germinable seeds ha<sup>-1</sup> with variants of wide-row sowing. It is proved that an increase in fertilizer rates ensured an increase in the share of seed fractions of 1.5 and 2.0 mm by 4.0-5.5 and 1.8-2.9%, respectively, while the share of seeds of 2.5 mm decreased by 2.0-3.7%.

The results of calculating the weight of 1,000 seeds are also important, as the matrix variability has weight expression and affects the level of individual seed productivity of plants (Makrushin, 1994). At a varietal idiotype of oilseed radish of the 'Zhuravka' variety with the index of mass of 1,000 seeds within 9-12 g, the studied technological approaches had essentially different efficiency of index formation including for seeds from pods of different zones. The average value of this indicator for the studied variants was 9.94 g for seeds from the pods of the lower zone and 9.33 g for seeds from the pods of the upper zone, which makes a difference of 6.5%. It should be noted that the difference in the value of the indicator differed within the technological options from 3.8% to 11.1%. Application of mineral fertilizers at a gradually increasing rate reduced the difference in weight of 1,000 seeds in comparison with seeds from pods of different zones by 1.3–3.7%. The maximum average difference in the value of the indicator for the two zones of pods was noted for the two extreme technological parameters in 4.0 and 0.5 million germinable seeds ha<sup>-1</sup> - 8.8 and 8.2%, respectively. As a result, the weight of 1,000 seeds had a stable growth dynamics with an increase in the feeding area of one plant against the background of fertilizer increase to N<sub>90</sub>P<sub>90</sub>K<sub>90</sub> with a maximum value for both zones on the level of 11.1–12.0 g and a minimum value in the range of 7.6–8.3 for the non-fertilized background of 4.0 million germinable seeds ha<sup>-1</sup>.

**Table 5.** Fraction composition of oilseed radish seeds by its separation on sieves with rectangular holes for the 'Zhuravka' variety on the brown pod phase (BBCH 85-87) in different tiers of inflorescence with different variants of agrophytocenosis formation on the average for 2013-2018, %

<u>2013–2010</u>	B	Fract	Fractions by seed separationon sieves with rectangular holes, mm (%)												
Sowingrate (C), sowing method	Sowingrate (C), sowing method	1.0×20	1.2×20	1.7×20	2.0×20	$2.2 \times 20$	2.5×20	3.0×20	1.0×20	$1.2 \times 20$	1.7×20	2.0×20	2.2×20	2.5×20	3.0×20
Sowin sowing	Sowin sowing		s fron rative	-	pods o	of the	upper	Seeds from the pods of the lower generative zone							
1	2	3	4	5	6	7	8	8	10	11	12	13	14	15	16
4.0	1	25.7	21.4	7.2	26.3	5.1	14.3	0.0	16.9	16.8	5.4	36.1	6.2	18.6	0.0
million,	2	25.0	21.9		25.2		14.8	0.0		17.4		35.2		18.7	0.0
row	3		22.9	7.9	24.7		15.6	0.0		17.8		34.7		19.6	0.0
2.0	4		23.4		24.2		16.2	0.0		18.1		35.6		20.1	0.0
3.0 million,	1 2	23.3	22.8 23.1	7.7	27.2 26.8		12.2 12.2	$\begin{array}{c} 0.0\\ 0.0\end{array}$		17.5 18.2	6.1	34.8 33.7	7.5	18.6 18.8	$\begin{array}{c} 0.0 \\ 0.0 \end{array}$
row	2 3	22.0	23.1			7.2 7.8	12.2	0.0	14.7	18.7	0.7 7.1	32.1	7.9 9.3	20.3	0.0
101	4		23.0		25.8		12.4	0.0		19.1	8.2	31.1		20.5	0.0
2.0	1	22.9	23.4	7.4	28.7	7.5	10.1	0.0	14.2	18.8	6.9	31.9		19.7	0.0
million,	2	21.5	23.9		27.4		11.2	0.0		19.1	7.3	32.2		20.2	0.0
row	3	20.1	24.6	8.5	26.5	8.3	12.0	0.0	10.2	19.8	8.2	31.5	9.4	20.9	0.0
	4	17.7	25.4	9.2	25.7	8.9	13.1	0.0	9.3	20.2	8.7	30.8	9.7	21.3	0.0
1.0	1	20.3	23.9	8.7	28.3	7.8	11.0	0.0	12.6	19.9	7.7	30.2		20.5	0.0
million,	2	19.2	24.2		27.8		11.5	0.0	11.4	20.4		28.6		21.2	0.2
row	3		24.8		27.4		11.2	0.0	10.8	20.6		27.1	10.3	21.8	0.3
20	4				$\frac{26.8}{28.5}$		11.8	0.0	9.1	20.9		26.7	10.7	22.5	0.5
2.0 million,	1 2	17.9 16.8	16.9 17.2	9.1 0.5	28.5 27.8		18.2 18.9	$\begin{array}{c} 0.0\\ 0.0\end{array}$	9.3 8.3	13.9 14.1	7.9 8.2	34.1 33.5	8.3 9.1	26.5 26.8	$\begin{array}{c} 0.0 \\ 0.0 \end{array}$
wide-row	23	15.7	17.2		27.8		20.8	0.0	6.6	14.1		33.2		20.8	0.0
wide-iow	4	14.2		10.1	26.2		20.8	0.0	4.9	14.7			10.2	27.6	0.0
1.5	1	16.3	16.6	10.4	27.4	7.8	21.5	0.0	8.9	12.6	8.4	32.6	8.9	28.4	0.2
million,	2	15.9	16.9	10.7		7.4	21.9	0.0	8.7		8.7	31.3		28.8	0.3
wide-row	3	14.7	17.5	11.3	26.8	5.9	23.8	0.0	6.4	13.9	8.9	30.9	6.9	32.7	0.3
	4	13.5	18.2	11.6	26.4	5.1	25.2	0.0	4.3	13.3	9.3	30.7	7.2	34.8	0.4
1.0	1		15.8				21.2		7.5	11.8		30.8		30.4	0.6
million,	2		15.3				22.7	0.5	7.2	12.4			10.5		0.8
wide-row	3		15.2				23.2	0.5	6.1	12.9			11.2		0.8
0.5	4		14.6				$\frac{23.7}{22.5}$	$\frac{0.7}{0.5}$	4.7	13.3 9.3			$\frac{11.8}{12.2}$		$\frac{0.8}{0.7}$
0.5 million,	1 2		14.8 14.6					0.5 0.7	7.4 5.7	9.5 9.5	9.8 10.6		12.3 13.5		0.7 0.9
wide-row	3		14.0					0.7	5.3				13.3		0.9
where now	4		13.9					0.8	3.4				15.2		1.0
$LSD_{05} \text{ A}^*$	-		0.32					0.0					0.35		
$LSD_{05}B$			0.18						0.11	0.15	0.08	0.28	0.20	0.23	
$LSD_{05}C$		0.25	0.26	0.15	0.32	0.12	0.24		0.15	0.21	0.12	0.39	0.28	0.33	
$LSD_{05}D$			0.26										0.28		
$LSD_{05}AB$			0.45										0.49		
$LSD_{05}AC$			0.63										0.69		
$LSD_{05}AD$		0.60	0.63	0.37	0.80	0.29	0.59		0.37	0.52	0.29	0.97	0.69	0.81	

Table 5 (continued)

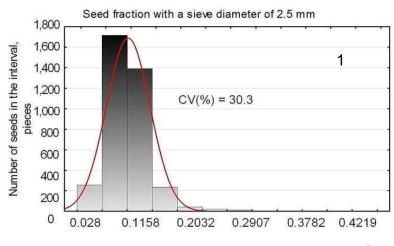
													(	
1	3	4	5	6	7	8	8	10	11	12	13	14	15	16
$LSD_{05}BC$	0.35	0.36	0.22	0.46	0.17	0.34		0.21	0.30	0.17	0.56	0.40	0.46	
$LSD_{05}BD$	0.35	0.36	0.22	0.46	0.17	0.34		0.21	0.30	0.17	0.56	0.40	0.46	
$LSD_{05}CD$	0.49	0.51	0.31	0.65	0.24	0.48		0.30	0.42	0.24	0.79	0.57	0.66	
$LSD_{05}ABC$	0.85	0.89	0.53	1.12	0.41	0.83		0.52	0.73	0.41	1.37	0.98	1.14	
$LSD_{05}ABD$	0.85	0.89	0.53	1.12	0.41	0.83		0.52	0.73	0.41	1.37	0.98	1.14	
LSD05ACD	1.21	1.26	0.75	1.59	0.58	1.18		0.74	1.04	0.58	1.93	1.39	1.61	
$LSD_{05}BCD$	0.70	0.73	0.43	0.92	0.33	0.68		0.43	0.60	0.34	1.12	0.80	0.93	
LSD05ABCD	1.71	1.78	1.06	2.25	0.82	1.66		1.04	1.47	0.83	2.73	1.96	2.28	
Share of influence	e													
А	20.4	22.3	12.3	55.01	22.8	13.8		8.74	21.4	1 30.21	57.43	320.33	323.03	
В	55.0	59.7	46.6	7.13	11.8	74.4		63.0	62.58	330.41	4.68	17.00	)65.64	
С	12.7	3.51	24.1	8.39	32.4	0.31		8.26	0.60	27.53	30.28	335.53	34.68	
D	9.12	3.25	2.13	2.78	4.64	3.01		16.1	1.75	7.93	3.54	6.45	3.11	
AB	0.85	0.95	0.59	0.19	0.14	0.99		0.95	1.00	0.36	0.14	0.21	0.85	
AC	0.19	0.06	0.35	0.16	0.55	0.03		0.11	0.03	0.46	0.50	0.56	0.10	
AD	0.10	0.06	0.06	0.05	0.13	0.08		0.19	0.07	0.20	0.06	0.15	0.10	
BC	0.17	4.25	13.5	25.22	216.5	6.29		1.27	12.00	51.43	0.60	8.62	1.16	
BD	0.60	0.21	0.01	0.02	3.14	0.40		0.05	0.05	0.24	0.17	0.48	0.19	
CD	0.41	3.98	0.06	0.29	3.63	0.26		0.63	0.02	0.47	1.61	4.81	0.75	
ABC	0.02	0.13	0.16	0.61	0.24	0.07		0.01	0.28	0.10	0.09	0.12	0.04	
ABD	0.01	0.01	0.00	0.00	0.04	0.01		0.00	0.00	0.00	0.01	0.01	0.01	
ACD	0.01	0.07	0.01	0.02	0.08	0.01		0.01	0.01	0.02	0.05	0.10	0.01	
BCD	0.36	1.47	0.03	0.10	3.86	0.32		0.54	0.13	0.61	0.81	5.54	0.33	
ABCD	0.01	0.03	0.00	0.02	0.05	0.01		0.01	0.01	0.02	0.04	0.08	0.02	

\* – in the system of value expression at deprivation of values,%; \*\* – 1 – Fertilizer-free;  $2 - N_{30}P_{30}K_{30}$ ;  $3 - N_{60}P_{60}K_{60}$ ;  $4 - N_{90}P_{90}K_{90}$ .

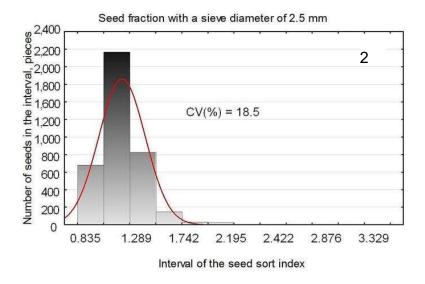
Under these conditions have also determined that the share of influence of the studied factors had different efficiency in the formation of values of different fractions of seeds separated on the round sieves, and within certain zones of pods placement. The most determinant factors in the formation of indicators of fractional composition of seeds were determined as the conditions of the year (factor A) with the share of influence in the range 10.3–67.9, sowing rate (factor C) 0.99–64.18. For the share of row width (factor B) and fertilizer (factor D) 5.0–23.26 and 0.2–47.41, respectively.

The certain non-uniformity of influence of the studied factors and their interaction indicates the complex mechanism of formation of linear and weight characteristics of seeds within the limits of fruit elements at different levels of combination of factors of the research. However, the regulatory factors of fractional composition of seeds and the expression of its matrix quality variability can be placed in the following order of importance A>C>D>B>BC. Similar results were obtained with respect to the quality variability of seeds in fractional composition within different zones of oilseed radish fruit elements at its separation on the sieves with rectangular holes (Table 5). The presented nature of fraction distribution has a wider interval nature, which indicates in favor of technologically restricted variant of oilseed radish seed separation on round sieves. This is indicated particularly by the corresponding share of seed fractions with the size of  $1.7 \times 20$  mm and  $2.2 \times 20$  mm. The nature of the fractional composition was different for seeds from pods of different zones.

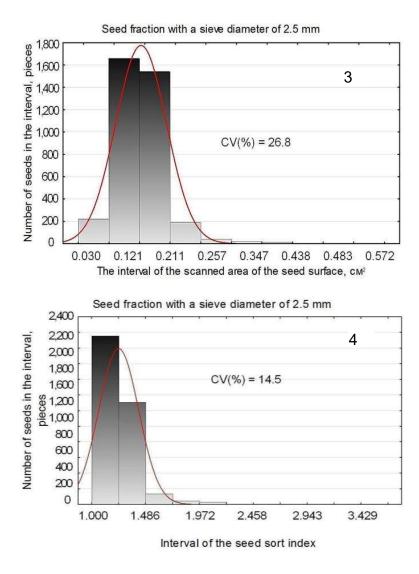
For the average variant on technological variants in seeds from pods of the lower zone of the generative part the share of fraction  $1.0 \times 20$  mm is 45.1% lower than for the variant from pods of the upper zone. For fractions  $1.2 \times 20$  and  $1.7 \times 20$ , respectively, it was 21.2% and 23.6% lower. However, the total increase in the share of fractions in the range from 2.0-2.5×20, was noted, respectively, by 21.7%, 13.3% and 45.5%. In addition, considering certain technological variants, the formation of fractional composition has the same features as in the case of separation on round sieves. These features include a consistent increase in the share of large seed fractions (2.2 and 2.5 mm) with an increase in a single plant's feeding area and an increase in background fertilizer. In addition, the nature of growth has two levels: one is observed in the range of normal line sowing. The other is observed in the range of wide-row sowing. Due to these reasons, the factor interaction of the research data system has a different nature of relationships among their values: B>A>C>D>BC. That is, the main components that determine the fractional distribution of seeds by separation on rectangular sieves is the width of row spacing and conditions of the year. It should also be noted that the sowing rate factor has two realizable components both as a single component and as its interaction with the width of row spacing.



The interval of the scanned area of the seed surface,  $\ensuremath{\mathsf{cm}}^2$ 



**Figure 8. (a)** Interval distribution of seeds of the 'Zhuravka' variety by indicators of the area of the scanned surface and sorting index of seeds for different fractions of seeds from the pods of the upper zone (position 1-2) of the generative part of plants with a technological variant of 2.0 million germinable seeds ha<sup>-1</sup> with fertilizer N90P90K90, 2013.



**Figure 8. (b).** The same from the pods of the lower zone (position 3–4), 2013.

On the other hand, it is important for practical seed production of cruciferous crops to understand the internal alignment of the received fractions of seeds (Würschum et al., 2012; Yang et al., 2016, 2017; Zheng et al., 2017). By applying the same electronic scanning approach to the separated seeds, we obtained its interval values of the morphological features of the seeds used to assess their variability (Table 3). So, for example, for the technological variant of 2.0 million germinable seeds ha<sup>-1</sup> with fertilizer N90P90K90 (conditions of 2016), the nature of the interval row of seeds by the size of the area of the scanned surface (SA) andsorting index (SI) for two fractions of 2.5 mm (round sieves) (Fig. 8 (a, b)) and  $2.5 \times 20$  (rectangular sieves) (Fig. 9) for different zone was different I also have found that the general variability of certain intervals for both given indicators on the value of the variation coefficient (CV) both for the fractions sieves is higher in the variant of seeds from the pods of the upper zone.

In particular, Figs 8, 9 gives an example of conditions for 2013 as approximate to the long-time average annual hydrothermal regime of vegetation of oilseed radish plants. Moreover, it should be noted that the difference in the variation coefficient between seeds from both pod zones was 3.5–9.7% higher for seeds from the upper zone. It was determined a higher value in the range of annual values for these deviations in the stress dry years (2015, 2017), and a lower value in the same range of values for conditions of years with moderate or optimal moistening (2013, 2014).

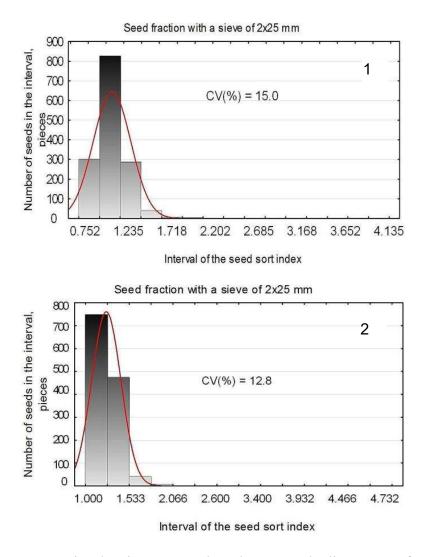


Figure 9. Interval distribution of seeds of the 'Zhuravka' variety by indicators of sorting index of seeds for seed fractions 2×25 mm from the pods of the upper zone (position 1) and from the pods of the lower zone (position 2) of the generative part of plants with a technological variant 2.0 million of germinable seeds ha<sup>-1</sup> with fertilizer N<sub>90</sub>P<sub>90</sub>K<sub>90</sub>, 2013.

It is also important that the general alignment of seeds from both zones of the generative part of plants is higher than the separation of seeds on rectangular sieves. For example, in Figs. 8 (a b) and 9, the coefficient of variation in the separation of seeds on round sieves was 2.2–5.4% higher, compared to a similar indicator for the separation of seeds on rectangular sieves.

#### CONCLUSION

Thus, on the basis of a comprehensive analysis of long-term data we have determined the presence of matrix quality variability of oilseed radish seeds both in morphological and weight characteristics according to the nature of placement of pods in the generative part of plants. The level of seed quality by matrix type in oilseed radish is determined by the nature of inflorescence formation and the corresponding morphotype of fruit elements, variable according to their height. The difference in the coefficient of variation of linear seed sizes was on average 6.7% higher for the upper zone of oilseed radish inflorescence compared to the lower zone. The maximum matrix variability of oilseed radish seeds was observed in the variant with a sowing rate of 0.5 million germinable seeds ha<sup>-1</sup> against the background of the application of 90 kg ha<sup>-1</sup> of mineral fertilizers. The integrated indicator of seed variability for this variant was the coefficient of 1.45 and 1.61 in comparison for the lower and upper inflorescence zones,

respectively, in the system of the ratio of the two variants 0.5 and 4.0 million germinable seeds ha<sup>-1</sup>. Mineral fertilizers contributed to the growth of seed variability in the range of 4.5-10.0% for the upper inflorescence zone depending on the cenosis design variant and 5.6-14.0% for the lower inflorescence zone. The heterogeneity of oilseed radish seed fractions in both round and elongated sieves was maximal at the minimum seeding rate on the background of maximum fertilizer. The summary result of the researchwas defined a decrease in the total linear individual size of oilseed radish seeds within the pod from the lower to the upper zone with a consistent reduction in the range of 3.8-8.3%, within the inflorescence in the range of 6.8-15.8%. An increase in the variation of seed morphoparameters in the comparison of the lower and upper zones of the inflorescence and pod by 7.2-10.2% with the maximum values of the growth of differences in the variants of the lowest seeding rate against the background of the maximum fertilizer N<sub>90</sub>P<sub>90</sub>K<sub>90</sub> was established.

#### REFERENCES

- Aicher, R.J., Larios, L. & Suding, K.N. 2011. Seed supply. recruitment. and assembly: quantifying relative seed and establishment limitation in a plant community context. *The American Naturalist* 178, 464–477.
- Al-Doori, S.A.M. & Hasan, M.Y. 2010. Effect of row spacing and nitrogen fertilization on growth, yield and quality of some rapeseed genotypes. *Journal Research* 9(4), 531–550.
- Ali, N., Javidfar, F. & Attary, A.A. 2002. Genetic variability.correlation and path analysis of yield and its components in winter rapeseed (*Brassica napus* L.). *Pakistan Journal of Botany* 34(2), 145–150.
- Alonso-Blanco, C., Aarts, M.G.M., Bentsink, L., Keurentjes, J.J.B., Reymond, M. & Vreugdenhil, D. 2009. What has natural variation taught us about plant development, physiology, and adaptation? *Plant Cell* 21, 1877–1896.
- Angadi, S.V., Cutforth, H.W., Miller, P.R., McConkey, B.G., Entz, M.H., Brandt, S.A. & Volkmar, K.M. 2000. Response of three Brassica species to high temperature stress during reproductive growth. *Canadian Journal of Plant Science* 80, 693–701.
- Balodis, O. & Gaile, Z. 2016. Sowing date and rate effect on winter oilseed rape (*Brassica napus* L.) yield components' formation. *Proceedings of the Latvian Academy of Sciences. Section* B. Natural, Exact, and Applied Sciences 70, 384–392.
- Batygina, T.B. 1999. Genetic heterogeneity of seeds: embryological aspects. *Plant Physiology* **46**(3), 438–454 (in Russian).
- Béreš, J., Bečka, D., Tomášek, J. & Vašák, J. 2019. Effect of autumn nitrogen fertilization on winter oilseed rape growth and yield parameters. *Plant, Soil and Environment* **65**(9), 435–441.
- Berry, P.M. & Spink, J.H. 2006. A physiological analysis of oilseed rape yields: Past and future. The *Journal of Agricultural Science* 144, 381–392.
- Berry, P.M., Spink, J.H., Foulkes, M.J. & White, P.J. 2010. The physiological basis of genotypic differences in nitrogen use efficiency in oilseed rape (*Brassica napus L.*). Field Crops Research 119, 365–373.
- Bouttier, C. & Morgan, D. 1992. Ovule development and determination of seed number per pod in oilseed rape (*Brassica napus* L.). *Journal of Experimental Botany* **43**, 709–714.
- Bukharov, A.F. & Baleev, D.N. 2012. The morphology of different quality seeds of vegetable umbrella crops. due to the place of formation on the mother plant. *Vegetable crops of Russia* 2(15), 44–47 (in Russian).
- Cavers, P. & Steel, M. 1984. Patterns of change in seed weight over time on individual plants. *The American Naturalist* **124**, 324–335.

- Chay, P. & Thurling, N. 1989. Variation in pod length in spring rape (*Brassica napus*) and its effect on seed yield and yield components. *Journal of agricultural science Cambridge* **113**, 139–147.
- Clarke, J.M. 1979. Intra-plant variation in number of seeds per pod and seed weight in *Brassica* napus. Tower. Canadian Journal of Plant Science **59**, 959–962.
- Donohue, K. & Schmitt, J. 1998. *Maternal environmental effects in plants: adaptive plasticity? In Maternal Effects as Adaptations* (Mousseau. T.A. & Fox.C.W. eds). Oxford University Press. 137–158.
- Dorofeyev, V.I. 2004. Fruits of the cruciferous: variety, structure, classification, origin. *Turczaninowia* 7(3), 76–87 (in Russian).
- Faraji, A. 2010. Flower formation and pod/flower ratio in canola (*Brassica napus* L.) affected by assimilates supply around flowering. *International Journal of Plant Production* **4**, 271–280.
- Faraji, A. 2012. Oil concentration in canola (*Brassica napus* L.) as a function of environmental conditions during seed filling period. *International Journal of Plant Production* **6**, 267–278.
- Fox, P.N., Crossa, J. & Ramagosa, I. 1997. *Multienvironment testing and genotype x environment interaction*. In: Kempton. R.A. and Fox P.N. (Eds.) Statistical methods for plant variety evaluation. Chapmann & Hall, pp. 117–138.
- Fu, Y., Wei, D., Dong, H., He, Y., Cui, Y., Mei, J. & Friedt, W. 2015. Comparative quantitative trait loci for silique length and seed weight in *Brassica napus*. *Scientific reports* **5**, 1–9.
- Gan, Y., Angadi, S.V., Cutforth, H., Potts, D., Angadi, V.V. & McDonald, C.L. 2004. Canola and mustard response to short periods of temperature and water stress at different developmental stages. *Canadian Journal of Plant Science* **84**, 697–704.
- Garcia, D., Fitz Gerald, J.N. & Berger, F. 2005. Maternal control of integument cell elongation and zygotic control of endosperm growth are coordinated to determine seed size in Arabidopsis. *Plant Cell* **17**, 52–60.
- Geritz, S.A., van der Meijden, E. & Metz, J.A. 1999. Evolutionary dynamics of seed size and seedling competitive ability. *Theoretical Population Biology* **55**, 324–343.
- Gattelmacher, B., Horst, W.J. & Becker, H.C. 1994. Factors that contribute to genetic variation for nutrient efficiency of crop plants. Journal of Plant Nutrition and Soil Science 157, 215–224.
- Gnan, S., Priest, A. & Kover, P.X. 2014. The genetic basis of natural variation in seed size and seed number and their trade-off using *Arabidopsis thaliana* MAGIC lines. *Genetics* **198**(4), 1751–1758.
- Gomez, N.V. & Miralles, D.J. 2011. Factors that modify early and late reproductive phases in oilseed rape (*Brassica napus* L.): its impact on seed yield and oil content. *Industrial Crops and Products* **34**, 1277–1285.
- Grant, C., Relf-Eckstein, J. & Zhou, R. 2013. *Impact of traditional and enhanced efficiency phosphorus fertilizers on canola emergence, yield, maturity and quality*. Canola Digest Science. Science Edition, 17 pp.
- Gunasekera, C.P., Martin, L.D., Siddique, K.H.M. & Walton, G.H. 2006. Genotype by environment interactions of Indian mustard (*Brassica juncea L.*) and canola (*Brassica napus L.*) in Mediteranneantype environments. 1. Crop growth and seed yield. *European Journal of Agronomy* 25, 1–12.
- Habekotté, B. 1996. Winter oilseed rape.analysis of yield formation and crop type design for higher yield potential. Ph.D. Thesis. Wageningen Agricultural University, Wageningen the Netherlands, 156 pp.
- Hasanuzzaman, M. 2020. The Plant Family Fabaceae. Biology and Physiological Responses to Environmental Stresses. Editors: Hasanuzzaman, M., Araújo, S. & Gill, S.S. (Eds.).
   Department of Agronomy Sher-e-Bangla Agricultural University Dhaka, Bangladesh, Springer Nature Singapore Pte Ltd., 531 pp.
- Hocking, D. 1997. Radish growing. NSW Agriculture Agfact 8(1), 32.

- Hocking, P.J., Kirkegarrd, J.A., Angus, J.F., Gibson, A.H. & Koetz, E.A. 1997. Comparison of canola, Indian mustard and linola in two contrasting environments. I. Effects of nitrogen fertilizer on dry-matter production, seed yield and seed quality. *Field Crops Research* 49, 107–125.
- Hocking, P.J. & Stapper, M. 2001. Effects of sowing time and nitrogen fertiliser on canola and wheat, and nitrogen fertiliser on Indian mustard. II. Nitrogen concentrations, N accumulation, and N fertiliser use efficiency. *Australian Journal of Agricultural Research* 52, 635–644.
- Horst, W., Behrens, T., Heuberger, H., Kamh, M., Reidenbach, G. & Wiesler, F. 2003. Genotypic differences in nitrogen use-efficiency in crop plants. In Innovative Soil-plant Systems for Sustainable Agricultural Production (Eds Lynch, J.M., Schepers, J.S. & Ünver, I.), Paris, France: OECD, pp. 75–92.
- Hua, W., Li, R.J., Zhan, G.M., Liu, J., Li, J. & Wang, X.F. 2012. Maternal control of seed oil content in *Brassica napus*: the role of silique wall photosynthesis. *The Plant Journal* **69**, 432–444.
- Ilyinska, A.P. 2013. Modern approaches to the classification of fruits *Brassicaceae* (on the example of flora of Ukraine). *Ukrainian Botanical Journal* **70**(4), 467–478 (in Ukrainian).
- Imbert, E. 2002. Ecological consequences and ontogeny of seed heteromorphism. *Perspectives in Plant Ecology, Evolution and Systematics* **5**(1), 13–36.
- Inanaga, S. & Kumura, A. 1987. Internal factors affecting seed set of rapeseed. In Seventh International Rapeseed Congress. Abstracts. *Polska angenja Interpress* **31**.
- IUSS Working Group WRB: World Reference Base for Soil Resources 2014. Update 2015. *World Soil Resources Reports* **106**, FAO, Rome, 203 pp.
- Ivanova, M.I. & Sarmosova, A.N. 2014. Comparative analysis of different quality seeds of cabbage cultures. *Bulletin of the Altai State Agrarian University* **4**(114), 5–9 (in Russian).
- Jankowski, K., Kijewski, Ł., Skwierawska, M., Krzebietke, S. & Mackiewicz-Walec, E. 2014. The effect of sulfur fertilization on the concentrations of copper, zinc and manganese in the roots, straw and cake of rapeseed (*Brassica napus* L. ssp. *oleifera* Metzg). *Journal of Elementology* 19, 433–446.
- Kaushik, N., Kumar, K., Kumar, S., Kaushik, N. & Roy, S. 2007. Genetic variability and divergence studies in seed traits and oil content of Jatropha (*Jatropha curcas* L.) accessions. *Biomass Bioenergy* 31, 497–502.
- Kennedy, Y., Yokoi, S., Sato, T., Daimon, H., Nishida, I. & Takahata, Y. 2011. Genetic variation of storage compounds and seed weight in rapeseed (*Brassica napus* L.) germplasms. *Breeding Science* **61**, 311–315.
- Khan, M.R.G., Ai, X.Y. & Zhang, J.Z. 2014. Genetic regulation of flowering time in annual and perennial plants. *Wiley Interdisciplinary Reviews: RNA* **5**(3), 347–359.
- Kindruk, N.A., Sechnyak, L.K. & Slyusarenko, O.K. 1990. *Environmental basics of seed production and forecasting of winter wheat seeds yield qualities*. Kyiv, Urozhay, 184 pp. (in Ukrainian).
- Kitaeva, I.E. 1952. *Quality varieties of turnip seeds cabbage rutabaga and radish and its causes:* authoreferat of Ph.D. Thesis. Moscow, 24 pp. (in Russian).
- Kizilova, E.G. 1974. Seed quality variability and its agronomic importance. Kiev: Urozhay, 216 pp. (in Russian).
- Kyrpa, N.Ya. 2011. *Principles and methods of seed mass separation*. In: Storage and processing of seeds **4**(142), 33–36 (in Ukrainian).
- Leishman, M.R., Westoby, M. & Jurado, E. 1995. Correlates of Seed Size Variation: A Comparison among Five Temperate Floras. *Journal of Ecology* **83**, 517–529.
- Lemontey, C., Mousset-Declas, C., Munier-Jolain, N. & Boutin, J.P. 2000. Maternal genotype influences pea seed size by controlling both mitotic activity during early embryogenesis and final endoreduplication level cotyledon cell size in mature seed. *Journal of Experimental Botany* **51**, 167–175.

Lester, W.Y., Wilen, R., Peta, W. & Bonham-Smith, C. 2004. High temperature stress of *Brassica napus* during owering reduces micro- and megagametophyte fertility. Induces fruit abortion.and disrupts seed production. *Journal of Experimental Botany* 55(396), 485–495.

Levina, R.E. 1987. Morphology and ecology of fruits. Leningrad, Nauka, 160 pp. (in Russian).

- Li, Y.P., Cheng, Y., Cai, G.Q., Fan, C.C. & Zhou, Y.M. 2014. Cytological basis and molecular mechanism of variation in number of seeds per pod in *Brassica napus*. Sci Sin Vitae 44, 822–831.
- Li, F., Chen, B., Xu, K., Wu, J., Song, W., Bancroft, I., Harper, A.L., Trick, M., Liu, S., Gao, G., Wang, N., Yan, G., Qiao, J., Li, J., Li, H., Xiao, X., Zhang, T. & Wu, X. 2014a. Genome-wide association study dissects the genetic architecture of seed weight and seed quality in rapeseed (*Brassica napus* L.). *DNA Research* 21, 355–367.
- Li, N. & Li, Y.H. 2015a. Maternal control of seed size in plants. *Journal of Experimental Botany* **66**, 1087–1097.
- Li, N., Peng, W., Shi, J.Q., Wang, X.F., Liu, G.H. & Wang, H.Z. 2015b. The natural variation of seed weight is mainly controlled by maternal genotype in rapeseed (*Brassica napus* L.). *PLOS ONE* 10:e0125360.
- Li, D., Lin, B G. & Zhang, D Q. 2016. Three-dimensional modeling of oilseed rape (*Brassica napus* L.) and its application in canopy architecture analysis. *Journal of Wuhan University of Technology Materials Science Edition* **62**, 575–580 (in Chinese).
- Li, N., Song, D., Peng, W., Zhan, J., Shi, J., Wang, X. & Liu, G. 2019. Maternal control of seed weight in rapeseed (*Brassica napus* L.): the causal link between the size of pod (mother, source) and seed (offspring, sink). *Plant Biotechnology Journal* 17, 736–749.
- Li, S., Zhu, Y., Varshney, R.K., Zhan, J., Zheng, X. & Shi, J. 2020. A systematic dissection of the mechanisms underlying the natural variation of silique number in rapeseed (*Brassica napus* L.) germplasm. *Plant Biotechnology Journal* 18, 568–580.
- Lotova, L.I. & Rudko, A.I. 1999. Anatomical features of fruits of different morphological types in the cruciferous family. *Bulletin of MOIP*. Biology section 104, **6**, 49–57.
- Lovett-Doust, J. 1989. Plant reproductive strategies and resource allocation. *Trends in Ecology & Evolution* **4**, 230–234.
- Luo, T., Zhang, J., Khan, M.N., Liu, J., Xu, Z. & Hu, L. 2018. Temperature variation caused by sowing dates significantly affects floral initiation and floral bud differentiation processes in rapeseed (*Brassica napus* L.). *Plant Science* **271**, 40–51.
- Makrushin, M.M. 1994. Seed science of field crops. Kiev: Urozhay. 208 pp. (in Ukrainian).
- Marjanovi'c-Jeromela, A., Nagl, N., Gvozdanovi'c-Varga, J., Hristov, N., Kondi'c-Špika, A., Vasi'c, M. & Marinkovi'c, R. 2011. Genotype by environment interaction for seed yield per plant in rapeseed using AMMI model. *Pesquisa Agropecuária Brasileira* **46**, 174–181.
- Mazer, S. & Wolfe, L. 1998. Density-mediated maternal effects on seed size in wild radish: genetic variation and its evolutionary implications. In Maternal Effects as Adaptations (Mousseau. T.A. and Fox.C.W..eds). Oxford University Press. pp. 323–343.
- McGregor, D.I. 1981. Pattern of flower and pod development in rapeseed. *Canadian Journal of Plant Science* **61**, 275–282.
- Meier, U. 2001. *BBCH Monograph*. Federal Biological Research Centre for Agriculture and Forestry. 2-th. Edition, 158 pp.
- Monty, A., Maebe, L., Mahy, G. & Brown, C.S. 2016. Diaspore heteromorphism in the invasive Bromus tectorum L. (Poaceae): sterile florets increase dispersal propensity and distance. Flora 224, 7–13.
- Morgan, D.G. 1980. Factors affecting fruit and seed development in field beans and oilseed rape. In Joint DPGRG and BPGRG Symposium Aspects and Prospects of Plant Growth Regulators. Monograph, 6. 151–164.
- Morrison, M.J. & Stewart, D.W. 2002. Heat stress during flowering in summer *Brassica*. *Crop Science* **42**, 797–803.

- Nikolaeva, M.H., Lyanguzova, I.V. & Pozdova, L.M. 1999. Seed biology. Saint Petersburg: Research Institute of chemistry, 232 pp. (in Russian).
- Olsson, G. 1960. Some relations between number of seeds per pod. seed size and oil content and the effects of selection for these characters *Brassica* and *Sinapsis*. *Hereditas* **46**, 27–70.
- Ozturk, O. 2010. Effects of source and rate of nitrogen fertilizer on yield, yield components and quality of winter rapeseed (*Brassica napus* L.). *Chilean Journal of Agricultural Research* **70**, 132–141.
- Pechan, P.M. 1988. Ovule fertilization and seed number per pod determination in oil seed rape (*Brassica napus*). *Annals of Botany* **61**, 201–207.
- Polowick, P.L. & Sawhney, V.K. 1986. A scanning electron microscopic study on the initiation and development of floral organs *Brassica napus* (cv. Westar). *American Journal of Botany* 73, 254–263.
- Polyovy, A.M. & Bozhko, L.Yu. 2007. *Long-term agro-meteorological forecasts*. Kyiv. KNT, 293 pp. (in Ukrainian).
- Porter, M.J., Pan, W.L., Schillinger, W.F., Madsen, I.J., Sowers, K.E. & Tao, H. 2020. Winter canola response to soil and fertilizer nitrogen in semiarid Mediterranean conditions. *Agronomy Journal* 112, 801–814.
- Puri, V. 1941. Studies in floral anatomy: I. Gynoecium constitution in the *Cruciferacea* Proc. *Indian Academy of Sciences* **41**, 166–187.
- Qian, P., Urton, R., Schoenau, J.J., King, T., Fatteicher, C. & Grant, C. 2012. Effect of seedplaced ammonium sulfate and monoammonium phosphate on germination, emergence and early plant biomass production of brassicae oilseed crops. In: Oilseeds. U.G. Akpan [Ed]. Rijeka: Intech Publishing Inc. pp. 53–62.
- Rabotnov, T.A. 1992. *Phytocoenology: Textbook for universities*. 3rd edition. Revisions and additions. Moscow, Publishing House of Moscow State University, 352 pp. (in Russian).
- Ragab, R., Battilani, A., Matovic, G., Stikic, R., Psarras, G. & Chartzoulakis, K. 2015. Saltmed model as an integrated management tool for water, crop, soil and N-fertilizer water management strategies and productivity: field and simulation study. *Irrigation and Drainage* 64, 13–28.
- Rahman, M., & McVetty, P.B.E. 2011. A review of Brassica seed color. *Canadian Journal of Plant Science* **91**, 437–446.
- Rao, G.U., Jain, A. & Shivanna, K.R. 1992. Effects of high temperature stress on Brassica pollen: viability. germination and ability to set fruits and seeds. *Annals of Botany* **69**, 193–198.
- Rathke, G.W., Christen, O. & Diepenbrock, W. 2005. Effects of nitrogen source and rate on productivity and quality of winter oilseed rape (*Brassica napus* L.) grown in different crop rotations. *Field Crops Research* 94, 103–113.
- Rees, M. 1996. Evolutionary ecology of seed dormancy and seed size. *Philosophical Transactions of the Royal Society B: Biological Sciences* **351**, 1299–1308.
- Roach, D.A. & Wulff, R.D. 1987. Maternal effects in plants. *Annual Review of Ecology, Evolution, and Systematics* 18, 209–235.
- Sadeghi, H., Khazaei, F., Sheidaei, S. & Yari, L 2011. Effect of seed size on seed germination behavior of safflower (*Carthamus tinctorius* L.). *Journal of Agriculture and Biological Sciences* 6, 5–8.
- Sadras, V.O., Rebetzke, G.J. & Edmeades, G.O. 2013. The phenotype and the components of phenotypic variance of crop traits. *Field Crops Research* **154**, 255–259.
- Sayko, V.F. 2011. *Features of research on cruciferous oil crops*. Kiev: 'Institute of Agriculture of the NAAS of Ukraine', 76 pp. (in Ukrainian).
- Scarisbrick, D.H., Daniels, R.W. & Noor Rawi, A.B. 1982. The effect of varying seed rate on the yield and yield components of oilseed rape (*Brassica napus*). Journal of Agricultural Science 99, 561–568.
- Schiessl, S.V., Qian, W., Iniguez-Luy, F. & Snowdon, R.J. 2015. Diverse regulatory factors associate with flowering time and yield responses in winter-type *Brassica napus*. *BMC Genomics* 16, 737.

- Skinner, R.H. & Moore, K. 2007. Chapter 4: Growth and Development of Forage Plants. In: Barnes, R.F., Moore, K.J., Nelson, C.J., Collins, M. editors. Forages. Volume 2: The Science of Grassland Agriculture. Sixth Edition. Ames, Iowa: Blackwell Publishing, pp. 53–66.
- Sieling, K., Böttcher, U. & Kage, H. 2017. Effect of Sowing Method and N Application on Seed Yield and N Use Efficiency of Winter Oilseed Rape. *Agronomy* 7, 21.
- Silveira, F.A.O., Negreiros, D., Araújo, L.M. & Fernandes, G.W. 2012. Does Seed Germination Contribute to Ecological Breadth and Geographic Range? A Test with Sympatric Diplusodon (Lythraceae) Species from Rupestrian Fields. Plant Species Biology 27, 170–173.
- Smith, L.J. & Scarisbrick, D.H. 1990. Reproductive development in oilseed rape (*Brassica napus* cv. Bienvenu). *Annals of Botany* **65**, 205–212.
- Stankowski, S., Bury, M., Jaroszewska, A, Michalska, B. & Gibczyńska, M. 2019. Effect of multi-component fertilizers on seeds yield, yield components and physiological parameters of winter oilseed rape (*Brassica napus* L.). Agronomy Research 17(5), 2071–2081.
- State standard of Ukraine 4138–2002. 2003. Seeds of agricultural plants. Methods of quality determination. Valid since 2004.01.01. Kiev, Derzhspozhyvstandart, 173 pp. (in Ukrainian).
- Sundaresan, V. 2005. Control of seed size in plants. *Proceedings of the National Academy of Sciences USA* **102**, 17887–17888.
- Susko, D.J. & Lovett-Doust, L. 2000. Patterns of seed mass variation and their effects on seedlings traits on *Alliaria petiolata (Brassicaceae)*. *American Journal of Botany* **87**(1), 56–66.
- Takashima, N.E., Rondanini, D.P., Puhl, L.E. & Miralles, D.L. 2013. Environmental factors affecting yield variability in spring and winter rapesed genotypes cultivated in the southeastern Argentine Pampas. *European Journal of Agronomy* **48**, 88–100.
- Tayo, T.O. & Morgan, D.G. 1978. Factors influencing flower and pod development in oilseed rape (*Brassica napus* L.). *The Journal of Agricultural Science Cambridge Core* **92**, 363–373.
- Test Guidelines for the conduct of tests for distinctness. uniformity and stability of Fodder Radish (Raphanus sativus L. var. oleiformis Pers.). Geneva. 2017. 19 pp.
- Tsytsiura Ya.H. 2018a. Features of layering formation in the oilseed radish agrophytocenosis under conditions of the right-bank forest-steppe of Ukraine. *Scientific bulletin of the National University of Life and Environmental Sciences of Ukraine*. Issue **286**. Series: Agronomy, pp. 205–215 (in Ukrainian).
- Tsytsiura, Ya.H. 2018b. Peculiarities of oilseed radish sowing desiccation considering the morphogenesis of its fruit elements under conditions of the the right-bank forest-steppe of Ukraine. *Tavria scientific bulletin: Academic periodical* **100**(2), 118–129 (in Ukrainian).
- Tsytsiura, Ya.H. & Tsytsiura, T.V. 2015. *Oilseed radish. Strategy of use and cultivation for fodder and seeds: monograph.* Vinnytsia: FOP Danylyuk V.H. 590 pp. (in Ukrainian).
- Tsytsiura, Y.H. 2019. Evaluation of the efficiency of oil radish agrofitocenosis construction by the factor of reproductive effort. *Bulgarian Journal of Agricultural Science* **25**(6), 1161–1174.
- Venable, D.L. 1985. The evolutionary ecology of seed heteromorphism. *American Naturalist* **126**(5), 577–595.
- Vovchenko, Yu.V. & Fursova, G.K. 2012. *Mustard: biology of yield formation and seed formation: monograph*. Kharkiv: Fort. 136 pp. (in Ukrainian).
- Voytenko, V.F. 1989. Heterocarpium (heterodisporia) in angiosperms: concept analysis, classification, terminology. *Ukrainian Botanical Journal* **74**(3), 281–297 (in Ukrainian).
- Vujaković, M., Marjanović-Jeromela, A., Jovičić, D.; Lečić, N., Marinković, R., Jakovljević, N.
  & Mehandžić-Stanišić, S. 2014. Effect of plant density on seed quality and yield of oilseed rape (*Brassica napus* L.). *Journal on Processing and Energy in Agriculture* 18, 73–76.
- Vyshnivsky, P. S. & Slisarchuk, M.V. 2014. Seeding qualities and yield properties of seeds of winter rapeseed depending on fractional composition of seed material. *Collection of* scientific works of the National Research Center 'Institute of Agriculture of the National Academy of Sciences' 3, 197–204.

- Wang, X.J., Mathieu, A., Courne'de, P.H., Allirand, J.M., Jullien, A., de Reffye, P. & Zhangm, B.G. 2011. Variability and regulation of the number of ovules seeds and pods according to assimilate availability in winter oilseed rape (*Brassica napus* L.). *Field Crops Research* 122(1), 60–69.
- Wilczewski, E., Harasimowicz-Hermann, G. & Lemańczyk, G. 2020. Effect of sowing method and density on the physical properties of the seed bed and oilseed rape yield. *Agronomy Research* **18**(2), 628–639.
- Wolfe, L.M. & Mazer, S.J. 2005. Patterns of phenotypic plasticity and their fitness consequences in wild radish (*Raphanus sativus: Brassicaceae*). *Journal of Plant Sciences* **166**, 631–640
- Wulff, R.D. 1986. Seed size variation in Desmodium paniculatum. I. Factors affecting seed size. *Journal of Ecology* **74**, 87–97.
- Würschum, T., Liu, W., Maurer, H.P., Abel, S. & Reif, J.C. 2012. Dissecting the genetic architecture of agronomic traits in multiple segregating populations in rapeseed (*Brassica napus* L.). *Theoretical and Applied Genetics* **124**, 153–161.
- Xing, N.L., Fan, C.H. & Zhou, Y.M. 2014. Parental selection of hybrid breeding based on maternal and paternal inheritance of traits in rapeseed (*Brassica napus* L.). *PLOS ONE* 9:e103165.
- Yang, Z. & Midmore, D.J. 2005. Modeling plant resource allocation and growth partitioning in responses to environmental heterogeneity. *Ecological Modelling* **181**, 59–77.
- Yang, Y., Shi, J., Wang, X., Liu, G. & Wang, H. 2016. Genetic architecture and mechanism of seed number per pod in rapeseed: elucidated through linkage and near-isogenic line analysis. *Scientific Reports* 6, 1–10.
- Yang, Y., Wang, Y., Shi, J., Zhan, J., Wang, X. & Liu, G. 2017. Genetic and cytological analyses of the natural variation of seed number per pod in Rapeseed (*Brassica napus* L.). Front. Plant Sci. 8, 1–14.
- Yizhyk M.K. 2000. *Agricultural seed production: Formation. structure and properties of seeds*. Kharkiv. **P. 1**, 103 pp. (in Ukrainian).
- Yizhyk, M.K. 2000a. *Agricultural seed production: Realizing the potential of seeds*. Kharkiv, **P. 2**, 117 pp. (in Ukrainian).
- Zar, J H. 1984. Biostatistical analysis. Englewood Cliffs. NJ, Prentice-Hall, 718 pp.
- Zaytsev, G.N. 1984. *Mathematical statistics in experimental botany*. Moscow, Nauka, 424 pp. (in Russian).
- Zhang, H., Berger, J.D. & Milroy, S.P. 2013. Genotype x environment interaction studies highlight the role of phenology in specific adaptation of canola (*Brassica napus* L.) to contrasting Mediterranean climates. *Field Crops Research* 144, 77–88.
- Zhang, Y., Zhang, D., Yu, H., Lin, B., Fu, Y. & Hua, S. 2016. Floral initiation in response to planting date reveals the key role of floral meristem differentiation prior to budding in canola (*Brassica napus* L.). *Frontiers in Plant Science* 7, 1369.
- Zheng, M., Peng, C., Liu, H., Tang, M., Yang, H., Li, X. & Liu, J. 2017. Genome-wide association study reveals candidate genes for control of plant height, branch initiation height and branch number in rapeseed (*Brassica napus* L.). *Frontiers in Plant Science* **8**, 1246.
- Zhu, J. 1996. Analytic methods for seed models with genotype-environment interactions. *Acta Genetica Sinica* 23, 56–68.
- Zhu, J. & Weir, B.S. 1994. Analysis of cytoplasmic and maternal effects I. A genetic model for diploid plant seeds and animals. *Theoretical and Applied Genetics* **89**, 153–159.
- Zlobin, Yu.A. 2009. *Popular ecology of plants: current status, points of growth*. Sumy: University. Book, 263 pp. (in Russian).
- Zohary, M. 1948. Carpological studies in *Cruciferae*. Palestine Journal of Botany. Jerusalem Series 4(8), 158–165.
- Zou, X., Guan, M. & Guan, C. 2020. Identification and evaluation of high nitrogen nutrition efficiency in rapeseed (*Brassica napus* L.) germplasm. *Oil Crop Science* **5**, 114–120.